



## Development of novel hydrogels by functionalization of sterculia gum for use in anti-ulcer drug delivery

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### ABSTRACT

In view of the pharmacological importance of sterculia gum and drug delivery devices based on hydrogels, the present study is an attempt to synthesize sterculia gum and poly(AAm) based hydrogels by using *N,N'*-MBAAM as crosslinker and ammonium persulfate as initiator. The polymeric networks were characterized by SEMs, FTIR and swelling studies. The release dynamics of model anti-ulcer drug (ranitidine hydrochloride) from the hydrogels has been studied for the evaluation of the release mechanism and diffusion coefficients. The values of the diffusion exponent '*n*' (0.43, 0.44 and 0.62) and gel characteristic constant '*k*' ( $8.669 \times 10^{-2}$ ,  $8.016 \times 10^{-2}$  and  $2.964 \times 10^{-2}$ ) have been obtained, respectively, in distilled water, pH 2.2 buffer and pH 7.4 buffer. The release of drug from the hydrogels occurred through Fickian diffusion mechanism in distilled water and in pH 2.2 buffer, and through non-Fickian diffusion mechanism in pH 7.4 buffers.

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### 1. Introduction

Natural gums are promising biodegradable, nontoxic, freely available and less expensive polymeric materials for use in drug delivery systems (Bhardwaj, Kanwar, Lal, & Gupta, 2000). However, these materials have certain drawbacks, like uncontrolled rate of hydration, thickening, drop in viscosity on storage, microbial contamination and require functionalization/modification to overcome these problems (Durso, 1980). The modification of polysaccharides to develop the hydrogels is a powerful tool, to control the interaction of the polymer with drugs, to enhance the load capability and to tailor the release profile of the drug (Nishimura et al., 1993). Grafting and crosslinking of vinyl monomers are the common practices to modify and to improve the functional properties of polysaccharides and forms the three-dimensional polymeric networks (hydrogels), those swell quickly by imbibing a large amount of water (Mostafa & Morsy, 2004). Hydrogels based on polysaccharides have attracted the considerable attention for use in drug delivery (Cascone et al., 2001; Chen, Jo, & Park, 1995). Amount and type of polysaccharide backbone, monomer and crosslinker concentration affect the swelling ratio of the hydrogels (Mundargi, Patil, Agnihotri, & Aminabhavi, 2007; Mundargi, Patil, & Aminabhavi, 2007; Zhang, Wang, Lu, Yang, & Yan, 2005). Lower crosslinker concentration results in the formation of very loosely crosslinked networks

which increase the swelling and decrease the gel strength (Das, Wadhwa, & Srivastava, 2006; Korsmeyer & Peppas, 1981; Zohuriaan-Mehr, Motazedi, Kabiri, & Ershad-Langroudi, 2005). The releasing medium such as simulated gastric fluid and simulated intestinal fluid affects the release pattern of the drugs from the formulations (Ji, Xu, & Wu, 2007). Chaurasia and coworkers (Chaurasia et al., 2006) have studied the in vitro release of methotrexate loaded crosslinked guar gum microspheres in phosphate-buffered saline, gastrointestinal fluid of different pH, and rat cecal content. Significant increase in drug release has been observed in medium containing rat cecal content (Das et al., 2006).

The use of natural gums such as xanthan gum (Verhoeven, Verwaet, & Remon, 2006), guar gum (George & Abraham, 2007), gum arabic (Chang, Leung, Lin, & Hsu, 2006), gellan gum (Agnihotri, Jawalkar, & Aminabhavi, 2006; Coviello, Alhaique, Dorigo, Matricardi, & Grassi, 2007) in drug delivery have been reported in the literature. Gum arabic has been investigated as sustained-release of drugs in tablets formulations and the release profile has further increased by coating the tablet with polyvinyl acetate and ethylene vinyl acetate (Baveja, Ranga Rao, & Arora, 1988). Due to gelling property of gum arabic, rate of release has been decreased as the amount of gum arabic in the pellets increased (Ray, Batra, Bhowmick, & Behra, 1994). The formulations coated with guar gum degraded in dissolution medium containing rat cecal contents and have affected the release dynamics (Krishnaiah, Muzib, Rao, Bhaskar, & Satyanarayana, 2003). Guar gum alone has not efficiently controlled the release dynamics of the drug release. Preparations

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of enteric coated matrix tablets with gums containing polymeric matrix formed by the grafting of methacrylic acid on to gums have showed enhanced release rates in the simulated colonic fluids because of the microbial degradation or polymer solubilization phenomena (Mundargi, Patil, & Agnihotri, et al., 2007; Mundargi, Patil, & Aminabhavi, 2007). The sterculia gum has been used to control the release of varying proportions of two model drugs, caffeine and diclofenac sodium, which have different solubility in aqueous medium (Munday & Cox, 2000).

Sterculia gum, is a medicinally important naturally occurring polysaccharide, composed of galacturonic acid, beta-D-galactose, glucuronic acid, L-rhamnose, and other residues (Weiping, 2000). It exudes from the tree *Sterculia urens*, belongs to the family 'Sterculiaceae' and is commonly known as karaya or sterculia etc. (Leung, 1980). Sterculia gum has unique features such as high swelling and water retention capacity, high viscosity, inherent nature of anti-microbial activity and abundant availability (Gauthami & Bhat, 1992). It has been used in the treatment of diarrhea (Huttel, 1983), ulcers (Love-Mignogna & Wind, 1978; May, 1982; Zide & Bevin, 1980), irritable bowel syndrome (Capron, Zeitoun, & Julien, 1981), chronic colonic diseases (Guerre & Neuman, 1979; Hunold, 1979) reducing cholesterol and improving glucose metabolism without adversely affecting most mineral balances (Behall, 1990; Behall, Scholfield, Lee, Powell, & Mores, 1987). Sterculia gum is an effective bulk laxative as the gum particles absorb water about 100 times of their original volume. In absorbing water the coarse particles swell enormously, forming a discontinuous type of mucilage that is very effective as a laxative (Meier, Seiler, & Stahelin, 1990; Verbeken, Dierckx, & Dewettinck, 2003). The swelling profile of sterculia gum has been used to produce denture adhesive and a bulk laxative (Park & Munday, 2004). In adhesive property, it has reduced 97 percent adhesion of *Streptococcus salivarius* to dentures in vivo (Wilson & Harvey, 1989) and has improved the adhesion in ileostomy and colostomy appliances (Kanamori et al., 2007).

In view of the pharmacological importance of sterculia gum and drug delivery devices based on hydrogels, sterculia gum, if suitably tailored to prepare the hydrogels, can act as the potential candidate for the novel drug delivery systems. Modification of the sterculia gum to develop the hydrogels is not much reported in the literature and the potential of sterculia gum to develop the drug delivery systems has not been explored. Therefore, the present study is an attempt, to synthesize sterculia gum and poly(AAm) based hydrogels by using *N,N'*-MBAAM as crosslinker and ammonium persulfate (APS) as initiator. The polymeric networks thus formed were named as sterculia-cl-poly(AAm) hydrogels thereafter, and have been characterized by FTIR and swelling studies. Swelling kinetics of the hydrogels has been studied as function of monomer concentration, initiator concentration, amount of sterculia and crosslinker concentration. Effect of pH and [NaCl] on swelling kinetics has also been studied. The release dynamics of model drug ranitidine hydrochloride from hydrogels has been studied for the evaluation of the release mechanism and diffusion coefficients.

## 2. Experimental

### 2.1. Materials and methods

Acrylamide (AAm) was obtained from Merck-Schuchardt, Germany. Ammonium persulfate (APS) was obtained from Qualigens Fine Chemicals Mumbai-India and *N,N'*-methylenebisacrylamide (*N,N'*-MBAAM) was obtained from Sisco Research Laboratory Pvt. Ltd. Mumbai, India. Ranitidine hydrochloride was obtained from the Scott-Edil Pharmacia Ltd. Himachal Pradesh, India. Sterculia gum was obtained from herbal medical store.

### 2.2. Synthesis of sterculia-cl-poly (AAm) polymers

Reaction was carried out with 1 g of sterculia gum, definite concentration of APS, definite concentration of monomer and crosslinker in 10 mL distilled water taken in test tube at 65 °C temperature for 2 h. The polymers thus formed were stirred for two hours in distilled water and for two hours in ethanol to remove the soluble fractions in the polymers and were then dried in oven at 40 °C. These polymers were named as [sterculia-cl-poly(AAm)]. The optimum reaction parameters were evaluated for the synthesis of sterculia-cl-poly(AAm) by varying [AAm] (from 0.281 to 1.407 mol/L), [APS] (from 4.386 to 21.930 mMol/L), amount of sterculia from 0.2 to 1.0 g and [*N,N'*-MBAAM] (from 6.486 to 32.432 mMol/L). These reaction parameters were evaluated on the basis of the swelling of the hydrogels after 24 h in distilled water, and shape and structural integrity maintained by the hydrogels during swelling. The optimum reaction conditions for the synthesis of sterculia-cl-poly(AAm) were obtained as [AAm] = 1.125 mol/L, [APS] = 13.158 mMol/L, sterculia gum = 0.8 g and [*N,N'*-MBAAM] = 6.486 mMol/L. Further sterculia-cl-poly(-AAm) hydrogels were synthesized at the optimum reaction conditions and were used to study the swelling kinetics hydrogels and release dynamics of drug from the hydrogels in different pH buffer.

### 2.3. Characterization

Sterculia gum and sterculia-cl-poly(AAm) polymers were characterized by the Scanning Electron Micrography (SEM), Fourier Transform Infrared Spectroscopy (FTIR) and swelling studies. To investigate and compare the surface morphology of sterculia gum and sterculia-cl-poly(AAm), SEMs were taken on ZEISS EVO 50 Microscope. FTIR spectra of sterculia and sterculia-cl-poly(AAm) polymers were recorded in KBr pellets on Nicolet 5700FTIR (THERMO).

Swelling of the polymeric networks was carried out in triplicate by gravimetric method. Known weight of polymers were taken and immersed in excess of distilled water for different time intervals at 37 °C and then polymers were removed, wiped with tissue paper to remove excess of solvent, and weighed immediately. The difference in weight gave the amount of water uptake by the polymers after definite time intervals. Swelling behavior of the polymer networks was studied as function of monomers concentration, initiator concentration, amount of sterculia and crosslinker concentration. Effect of pH and [NaCl] on swelling kinetics was also studied.

### 2.4. Release dynamics of the model drug

#### 2.4.1. Preparation calibration curves

In this procedure, the absorbance of a number of standard solutions of the reference substance at concentrations encompassing the sample concentrations were measured on the UV-Visible Spectrophotometer (Cary 100 Bio, Varian) at  $\lambda_{\max}$  313 nm and calibration graph was constructed. The concentration of the drug (ranitidine hydrochloride) in the sample solution was read from the graph as the concentration corresponding to the absorbance of the solution. Three calibration graphs were made in distilled water, pH 2.2 buffer and pH 7.4 buffer to determine the amount of drug release from the drug-loaded polymeric matrix in these medium.

#### 2.4.2. Drug loading to the polymer matrix

The loading of a drug onto hydrogels was carried out by swelling equilibrium method. The hydrogels were allowed to swell in the drug solution of known concentration for 24 h at 37 °C and then dried to obtain the release device (Singh, 2007).

#### 2.4.3. Drug release from polymer matrix

In vitro release studies of the drug were carried out by placing dried and loaded sample in definite volume of releasing medium at 37 °C temperature. The amount of drug released was measured spectrophotometrically in distilled water, pH 2.2 buffer and pH 7.4 buffer after every 30 min in each case (Singh, 2007).

#### 2.4.4. Preparation of buffer solution

Buffer solution of pH 2.2 was prepared by taking 50 mL of 0.2 M KCl and 7.8 mL of 0.2 N HCl in volumetric flask to make volume 200 mL with distilled water. Buffer solution of pH 7.4 was prepared by taking 50 mL of 0.2 M KH<sub>2</sub>PO<sub>4</sub> and 39.1 mL of 0.2 N NaOH in volumetric flask to make volume 200 mL with distilled water (Pharmacopoeia of India, 1985).

#### 2.5. Mechanism and mathematical modeling of drug release from polymer matrix

Based on the relative rate of diffusion of water into polymer matrix and rate of polymer chain relaxation, swelling of the polymers and the drug release profiles from the swelling polymers have been classified into three types of diffusion mechanisms (Alfrey, Gurnee, & Lloyd, 1966; Peppas & Korsmeyer, 1987). These mechanisms are Case I or Simple Fickian Diffusion, Case II Diffusion and Non-Fickian or Anomalous Diffusion (Ritger & Peppas, 1987a; Ritger & Peppas, 1987b). In the case of water uptake, the weight gain,  $M_s$ , is described by the Eq. (1)

$$M_s = kt^n \quad (1)$$

where  $k$  and  $n$  are constants. Normal Fickian diffusion is characterized by  $n = 0.5$ , while Case II diffusion by  $n = 1.0$ . A value of  $n$  between 0.5 and 1.0 indicates a mixture of Fickian and Case II diffusion, which is usually called non-Fickian or anomalous. Ritger

and Peppas have showed that the above power law expression could be used for the evaluation of drug release from swellable systems. In this case,  $M_t/M_\infty$  replace  $M_s$  in above equation to give Eq. (2). For cylindrical shaped hydrogels, the initial diffusion coefficients ( $D_i$ ), average diffusion coefficient  $D_A$  and late diffusion coefficients have been calculated from the Eqs. (3)–(5), respectively.

$$\frac{M_t}{M_\infty} = kt^n \quad (2)$$

$$\frac{M_t}{M_\infty} = 4 \left( \frac{Dt}{\pi \ell^2} \right)^{0.5} \quad (3)$$

$$D_A = \frac{0.049 \ell^2}{t^{1/2}} \quad (4)$$

$$\frac{M_t}{M_\infty} = 1 - \left( \frac{8}{\pi^2} \right) \exp \left[ \frac{(-\pi^2 Dt)}{\ell^2} \right] \quad (5)$$

where  $M_t/M_\infty$  is the fractional release of drug in time  $t$ , ' $k$ ' is the constant characteristic of the drug-polymer system, and ' $n$ ' is the diffusion exponent characteristic of the release mechanism.  $M_t$  and  $M_\infty$  is drug released at time ' $t$ ' and at equilibrium, respectively,  $D$  is the initial diffusion coefficient and ' $\ell$ ' is the thickness of the sample.  $t^{1/2}$  is the time required for 50% release of drug (Ritger & Peppas, 1987a; Ritger & Peppas, 1987b). The values of diffusion coefficients have been evaluated for the swelling of the polymer and for the release of the drug from the polymer and results are presented in Tables 1 and 2.

### 3. Results and discussion

#### 3.1. Characterization

The morphology of sterculia and sterculia-*cl*-poly(AAm) was examined by SEMs and is presented in the Fig. 1a and b, respec-

**Table 1**

Results of diffusion exponent ' $n$ ', gel characteristic constant ' $k$ ' and various diffusion coefficients for the swelling kinetics of sterculia-*cl*-poly(AAm) hydrogels in different medium at 37 °C

S. No.	Parameter	Diffusion exponent 'n'	Gel characteristic constant 'k' × 10 <sup>2</sup>	Diffusion coefficients (Cm <sup>2</sup> min <sup>-1</sup> )		
				Initial D <sub>i</sub> × 10 <sup>4</sup>	Average D <sub>A</sub> × 10 <sup>4</sup>	Late time D <sub>L</sub> × 10 <sup>4</sup>
Effect of [AAm]						
1	0.281 M/L	0.50	3.357	1.994	3.864	0.340
2	0.563 M/L	0.50	3.412	2.047	3.992	0.348
3	0.844 M/L	0.50	3.483	3.533	7.671	0.620
4	1.125 M/L	0.50	3.504	8.014	15.263	1.369
5	1.407 M/L	0.47	3.793	8.837	17.069	1.445
Effect of [APS]						
1	4.386 mM/L	0.50	3.504	8.014	15.263	1.369
2	8.772 mM/L	0.56	2.014	9.378	18.077	1.563
3	13.158 mM/L	0.57	2.042	10.857	19.746	1.757
4	17.544 mM/L	0.56	2.075	8.444	15.850	1.395
5	21.930 mM/L	0.54	2.845	10.289	17.555	1.662
Effect of amount of sterculia						
1	0.2 g	0.53	2.198	4.635	10.555	0.853
2	0.4 g	0.52	2.773	8.385	16.895	1.454
3	0.6 g	0.50	3.342	7.900	16.657	1.398
4	0.8 g	0.50	3.516	9.318	20.588	1.693
5	1.0 g	0.57	2.042	10.857	19.746	1.757
Effect of [N,N'-MBAAm]						
1	6.486 mM/L	0.50	3.516	9.318	20.588	1.693
2	12.973 mM/L	0.56	2.173	13.404	25.087	2.211
3	19.459 mM/L	0.53	2.655	12.926	24.629	2.168
4	25.945 mM/L	0.53	2.410	10.555	22.159	1.852
5	32.432 mM/L	0.60	2.061	16.707	25.771	2.545
Effect of pH						
1	Distilled water	0.50	3.516	9.318	20.588	1.693
2	pH 2.2 buffer	0.50	2.624	8.456	19.494	1.571
3	pH 7.4 buffer	0.50	2.904	8.736	20.323	1.618
Effect of salt						
1	Distilled water	0.50	3.516	9.318	20.588	1.693
2	0.9% NaCl	0.50	2.754	8.903	20.211	1.637



**Table 2**  
Results of diffusion exponent 'n', gel characteristic constant 'k' and various diffusion coefficients for the release of ranitidine hydrochloride from drug-loaded sterculia-*cl*-poly(AAm) hydrogels in different medium at 37 °C

S. No.	Drug releasing medium	Diffusion exponent 'n'	Gel characteristic constant 'k' × 10 <sub>2</sub>	Diffusion coefficients (Cm <sup>-2</sup> Min <sup>-1</sup> )		
				Initial $D_i \times 10^4$	Average $D_A \times 10^4$	Late time $D_L \times 10^4$
1	Distilled water	0.43	8.669	25.857	5.585	6.627
2	pH 2.2 buffer	0.44	8.016	30.691	5.760	7.576
3	pH 7.4 buffer	0.62	2.964	47.499	4.520	8.646

tively. It is observed from the SEMs that sterculia has smooth and homogeneous morphology whereas modified sterculia has structural heterogeneity.

FTIR spectra of sterculia and sterculia-*cl*-poly(AAm) were recorded to study the modification of the sterculia and are presented in Fig. 2a and b, respectively. The broad absorption bands at 3427.5 cm<sup>-1</sup> was observed due to —OH stretching which indicated association in the polymeric networks. Absorption bands due to C=O stretching of amide has been observed at 1670.1 cm<sup>-1</sup> in sterculia-*cl*-poly(AAm) apart from the usual peaks in sterculia gum.

### 3.2. Swelling kinetics of sterculia-*cl*-poly(AAm) hydrogels

Swelling parameters are the most important factor for hydrogel characterization, because a fundamental relationship exists between the swelling of a polymer and the nature of the swelling medium. In order to evaluate the optimum reaction parameters for the synthesis of sterculia-*cl*-poly(AAm) hydrogels, swelling of the hydrogels and structural integrity maintained by the hydrogels after 24 h swelling were taken as the criteria. Swelling of the poly-

mers was studied as a function of [AAm], [APS], amount of sterculia, [N,N'-MBAAm] in the polymer matrix and as a function of pH and [NaCl] of the swelling medium.

#### 3.2.1. Swelling as a function of monomer concentration

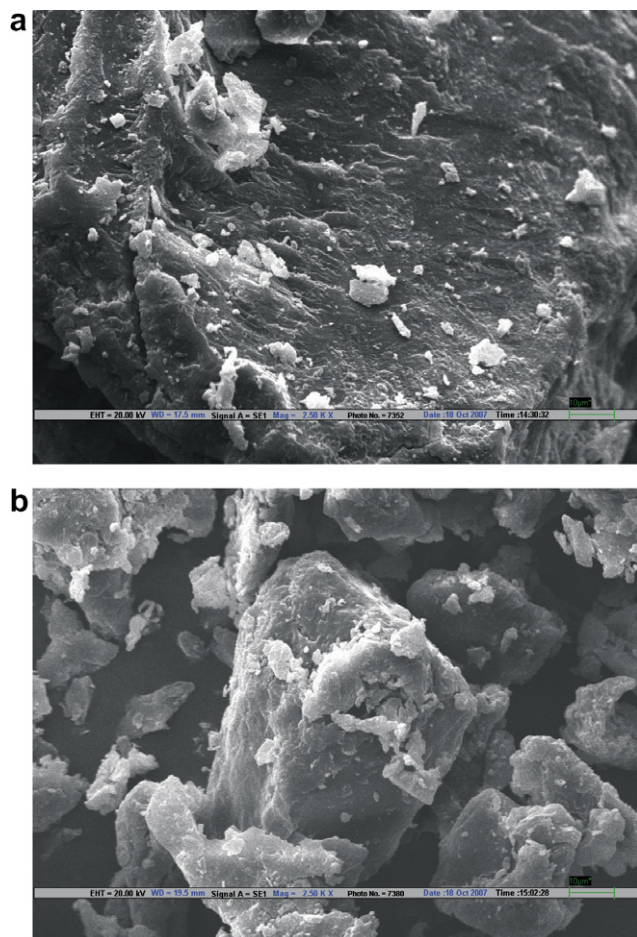
Effect of monomer concentration, used during the synthesis of hydrogels, on the swelling of the crosslinked polymers was studied as a function of [AAm] and results are presented in the Fig. 3.1. It has been observed from figure that swelling of polymer decreases with increase in monomer concentration in the polymer matrix. After 24 h, maximum (16.04 ± 2.05 g) water uptake has occurred by per gram of the polymer prepared with 0.281 mol/L of [AAm] but structural integrity has not been maintained by the hydrogels after 24 h swelling. Therefore, in the present case optimum concentration for the further synthesis of polymers was taken as 1.125 mol/L. At this concentration maximum water uptake (12.20 ± 1.46 g) has occurred after 24 h swelling and hydrogels have maintained its cylindrical shape. The values of diffusion exponent 'n' and gel characteristic constant 'k' have been evaluated from the slope and the intercept of the plot  $\ln M_t/M_\infty$  versus  $\ln t$  and the results are presented in the Table 1. It is clear from the values of the 'n' that the Fickian diffusion mechanism has occurred for the hydrogels prepared with different monomer concentration. The values of diffusion coefficients are presented in the Table 1.

#### 3.2.2. Swelling as a function of initiator

At the optimum monomer concentration, to observe the effect of initiator concentration on the network formation, the hydrogels were prepared with different initiator concentration. The [APS] was varied from 4.386 to 21.930 mMol/L and effect of this variation on the swelling of sterculia-*cl*-poly(AAm) hydrogels is presented in Fig. 3.2. Swelling of the polymer matrix decreases as the concentration of initiator increased during the synthesis of polymeric networks. This may be due to the reason that higher concentration of the initiator has started the formation of number of polymer chains which has decreased the chain length and affected the network formation. The maximum (13.25 ± 0.52 g) water uptake has occurred for the hydrogels prepared with 13.158 mMol/L of [APS]. It is clear from the values of the diffusion exponent that diffusion of water molecule from the polymer matrix occurred through non-Fickian type diffusion mechanism. The values of the initial and average diffusion coefficients have been obtained higher than the late diffusion coefficients which indicate that in the starting the rate of swelling of polymer matrix was higher than the latter stages of swelling.

#### 3.2.3. Swelling as a function of amount of sterculia gum

In order to study the effect of amount of sterculia gum on the structure of the polymer, the hydrogels were prepared with different gum contents and their swelling was taken in distilled water at 37 °C. Polymers were synthesized by varying the of sterculia gum from 0.2 to 1.0 g and results are presented in Fig. 3.3. The swelling of the polymers increases with the increase in sterculia contents in the composition of polymer matrix. This is probably due to the reason that higher degree of gum hydration has occurred in the hydrogels prepared with higher gum contents, which has increased the



**Fig. 1.** (Singh and Sharma) Scanning electron micrograph of (a) sterculia gum (b) sterculia-*cl*-poly(AAm).

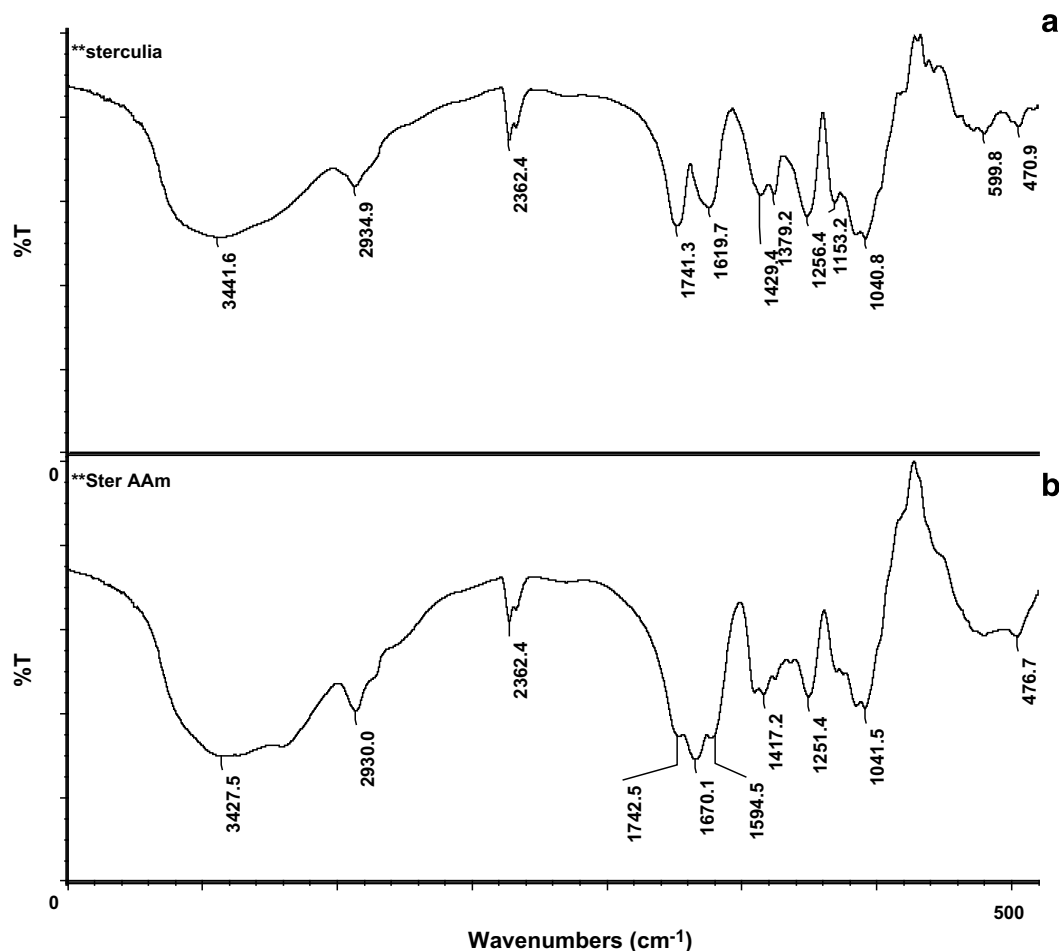


Fig. 2. (Singh and Sharma) FTIR spectra of (a) sterculia gum (b) sterculia-cl-poly(AAm).

number of intimate contacts between particle of gum and water, and led to high swelling (Patel & Patel, 2007). Maximum ( $12.62 \pm 0.94$  g) water uptake has occurred in the matrix prepared with 0.8 g of sterculia gum. The diffusion of water from the polymer matrix occurred through Fickian diffusion mechanism.

### 3.2.4. Swelling as a function of $[N,N'$ -MBAAm]

At the optimum concentration of monomer, initiator and sterculia gum, the polymers were prepared with different crosslinker concentration. The polymer networks were prepared by varying the crosslinker concentration from 6.486 to 32.432 mMol/L. Swelling of the sterculia-cl-poly(AAm) was studied as a function of  $[N,N'$ -MBAAm] at 37 °C and the results are presented in the Fig. 3.4. It has been observed from the figure that swelling decreases with the increase in the concentration of crosslinker in the polymer matrix. This is due to the fact that with the increase in the concentration of crosslinker, crosslinking density increases which decreases the pore size in the crosslinked matrix and thereafter decreases the swelling in the polymer. Maximum amount of water uptake ( $12.62 \pm 0.94$ ) per gram of gel has occurred in case of polymer prepared with 6.486 mMol/L of  $[N,N'$ -MBAAm]. The values of diffusion exponent 'n' is greater than 0.5, which indicate a non-Fickian type of diffusion mechanism occurred for the diffusion of water molecules in the polymer matrix prepared with different crosslinker concentration. The values of the diffusion coefficients for the swelling of the hydrogels prepared as a function of  $[N,N'$ -MBAAm] are presented in Table 1. It has been observed from the table that the values obtained for initial and average diffusion coefficient are higher than the late diffusion coefficient.

### 3.2.5. Swelling as a function of pH and $[NaCl]$ of the swelling medium

At the optimum reaction conditions, polymers were synthesized and were used to study the effect of reaction medium on the swelling mechanism. The swelling of polymers in different

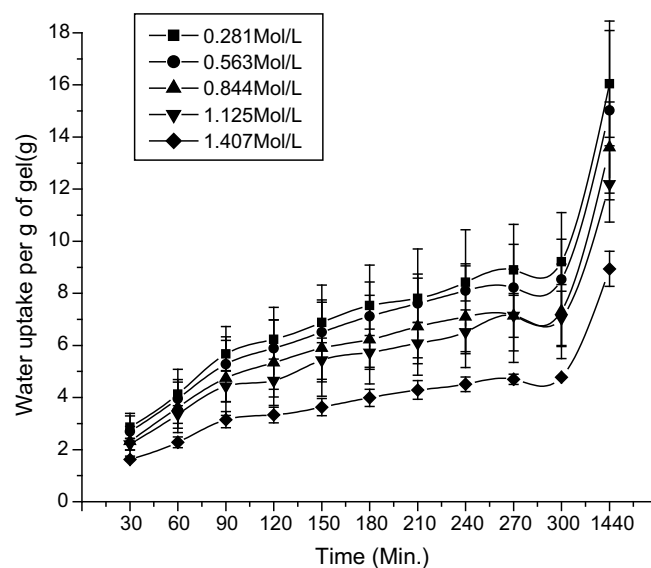
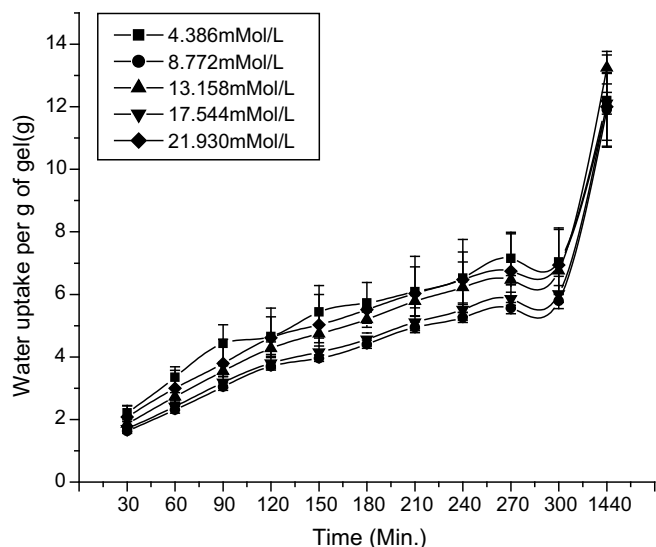
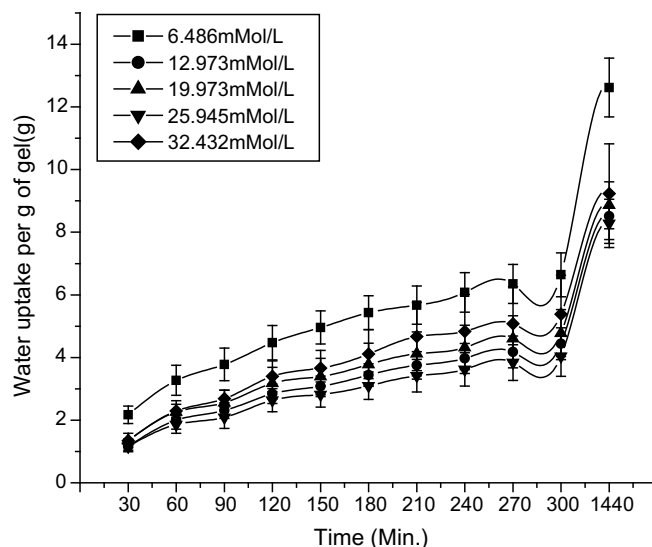


Fig. 3.1. (Singh and Sharma) Effect of [AAm] on swelling kinetics of sterculia-cl-poly(AAm) hydrogels in distilled water at 37 °C. {Sterculia gum = 1 g, [AP-S] = 4.386 mMol/L,  $[N,N'$ -MBAAm] = 6.486 mMol/L}.



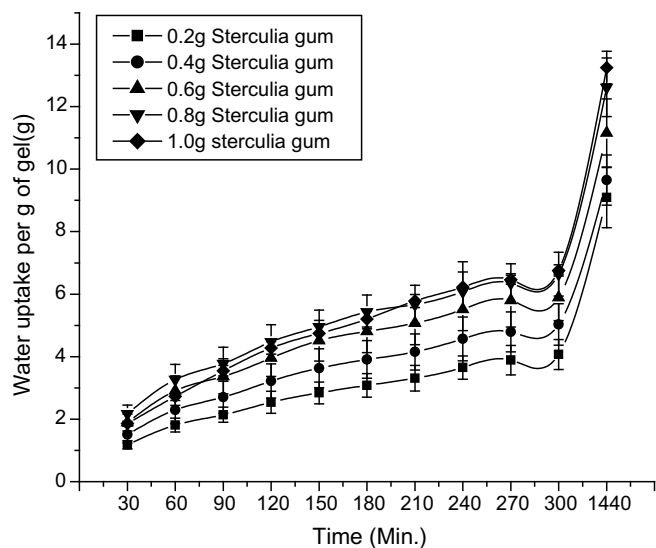
**Fig. 3.2.** (Singh and Sharma) Effect of [APS] on swelling kinetics of sterculia-cl-poly(AAm) hydrogels in distilled water at 37 °C. {Sterculia gum = 1 g, [AAm] = 1.125 Mol/L, [N,N'-MBAAm] = 6.486 mMol/L}.



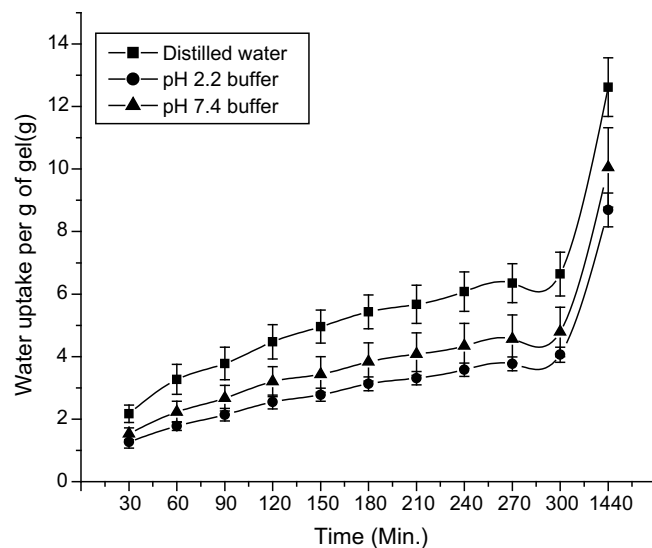
**Fig. 3.4.** (Singh and Sharma) Effect of [N,N'-MBAAm] on swelling kinetics of sterculia-cl-poly(AAm) hydrogels in distilled water at 37 °C. {Sterculia gum = 0.8 g, [APS] = 13.158 mMol/L, [AAm] = 1.125 Mol/L}.

pH and salt solution are presented in Figs. 3.5 and 3.6, respectively. When the effect of pH was studied the swelling was carried out in distilled water, pH 2.2 buffer, and pH 7.4 buffer for 24 h at 37 °C. It has been observed from the figure that swelling increases with increase in pH of the swelling medium and maximum ( $12.62 \pm 0.94$  g), ( $8.69 \pm 0.54$  g) and ( $10.05 \pm 1.27$  g) water uptake per gram of gel has occurred in distilled water, pH 2.2 buffer, pH 7.4 buffer, respectively. At lower pH, the  $-\text{CONH}_2$  groups do not ionize and keep the network at its collapse state. At high pH solution, these groups get partially ionize, and the charged  $-\text{COO}-$  groups repel each other and cause more swelling of the polymer. To study the effect of salt concentration, swelling of the hydrogels was carried out in 0.9% NaCl at 37 °C and amount of water uptake by per gram of gel in salt solution was observed less as compared to the distilled water. In distilled water and 0.9% [NaCl] per gram of the gel has taken maximum ( $12.62 \pm 0.94$  g) and ( $8.63 \pm 1.08$  g) water, respectively.

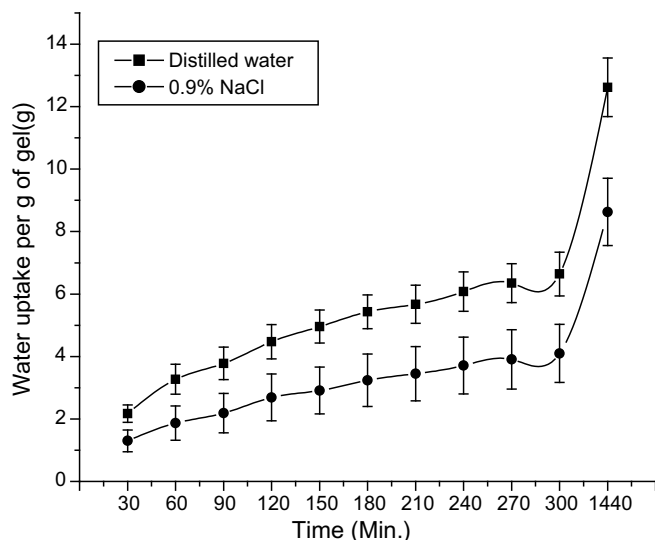
The values of the diffusion exponent ' $n$ ' has been obtained 0.5 in all the three mediums i.e. in distilled water, pH 2.2 buffer, and pH 7.4 buffer. These values show that swelling of the hydrogels occurred through Fickian type mechanism all the three releasing mediums. The values of the gel characteristic constant are different in different swelling medium which indicate that the polymer matrix showing different behavior in these mediums. In salt solution, hydrogel shows Fickian diffusion mechanism. In Fickian type diffusion mechanism the rate of diffusion of water molecules into the polymer matrix is much less than that of rate of relaxation of polymeric chains. The values of the initial diffusion coefficient, average diffusion coefficient and late diffusion coefficients for the swelling of sterculia-cl-poly(AAm) are presented in Table 1. In each swelling medium values of the initial and average diffusion coefficient have been observed higher than the other coefficients. It means the ini-



**Fig. 3.3.** (Singh and Sharma) Effect of amount of sterculia gum on swelling kinetics of sterculia-cl-poly(AAm) hydrogels in distilled water at 37 °C. {[AAm] = 1.125 Mol/L, [APS] = 13.158 mMol/L, [N,N'-MBAAm] = 6.486 mMol/L}.



**Fig. 3.5.** (Singh and Sharma) Effect of pH on swelling kinetics of sterculia-cl-poly(AAm) hydrogels at 37 °C. {Sterculia gum = 0.8 g, [AAm] = 1.125 Mol/L, [APS] = 13.158 mMol/L, [N,N'-MBAAm] = 6.486 mMol/L}.

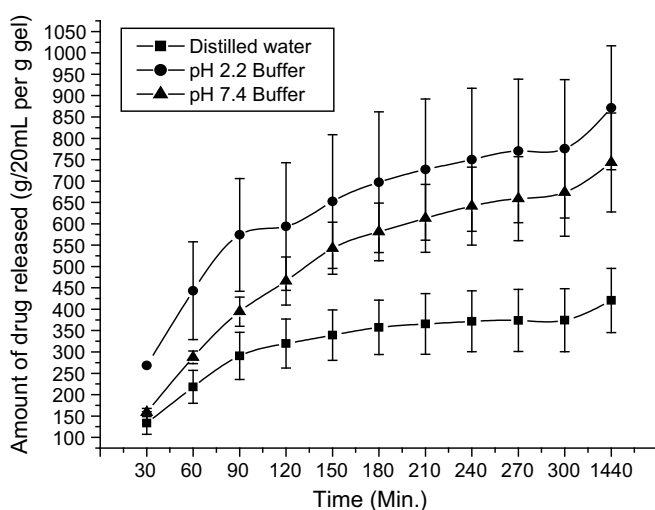


**Fig. 3.6.** (Singh and Sharma) Effect of [NaCl] on swelling kinetics of sterculia-cl-poly(AAm) hydrogels at 37 °C. {Sterculia gum = 0.8 g, [AAm] = 1.125 Mol/L, [APS] = 13.158 mMol/L, [N,N'-MBAAm] = 6.486 mMol/L}.

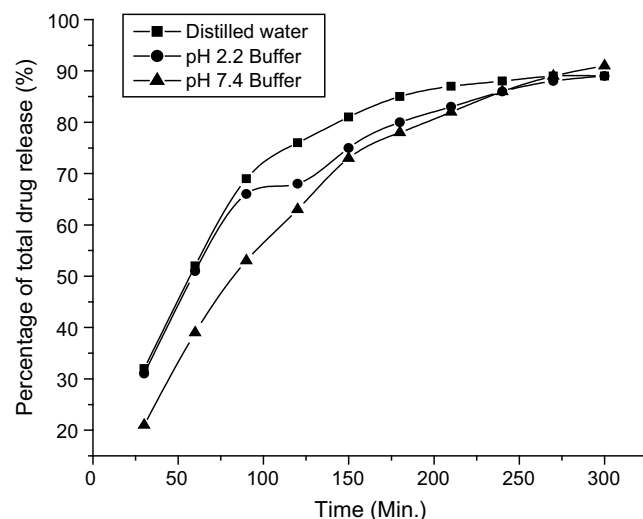
tial rate of diffusion of water molecule in the polymer matrix was higher than the late stages.

### 3.3. In vitro release dynamics of ranitidine hydrochloride

In vitro release profile of ranitidine hydrochloride from per gram of the drug-loaded hydrogel was studied in distilled water, pH 2.2 buffer and pH 7.4 buffer and results are presented in Fig. 4.1. It has been observed from the figure that the amount of drug released from per gram of the gel is higher in pH 2.2 solution as compared to the release of drug in solution of pH 7.4 and in distilled water. The trends obtained are not corresponding to the swelling pattern of the sterculia-cl-poly(AAm) hydrogels where the swelling has been observed higher in distilled water and pH 7.4 buffer. This may be due to the more solubility of ranitidine hydrochloride in pH 2.2 buffer. Fifty percent of the total release



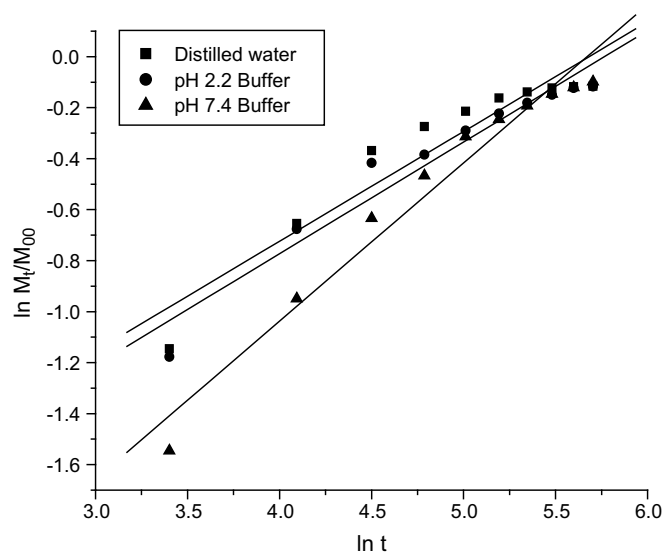
**Fig. 4.1.** (Singh and Sharma) Release profile of ranitidine hydrochloride from drug-loaded sterculia-cl-poly(AAm) hydrogels in different medium at 37 °C. {Sterculia gum = 0.8 g, [AAm] = 1.125 Mol/L, [APS] = 13.158 mMol/L, [N,N'-MBAAm] = 6.486 mMol/L}.



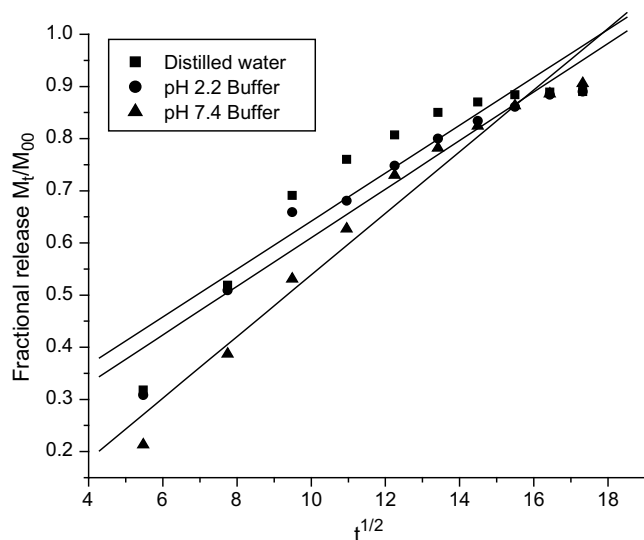
**Fig. 4.2.** (Singh and Sharma) Percentage of the total release of ranitidine hydrochloride from drug-loaded sterculia-cl-poly(AAm) hydrogels in different medium at 37 °C.

of the drug occurred in 47, 59 and 88 min in distilled water, pH 2.2 buffer and in pH 7.4 buffer, respectively (Fig. 4.2).

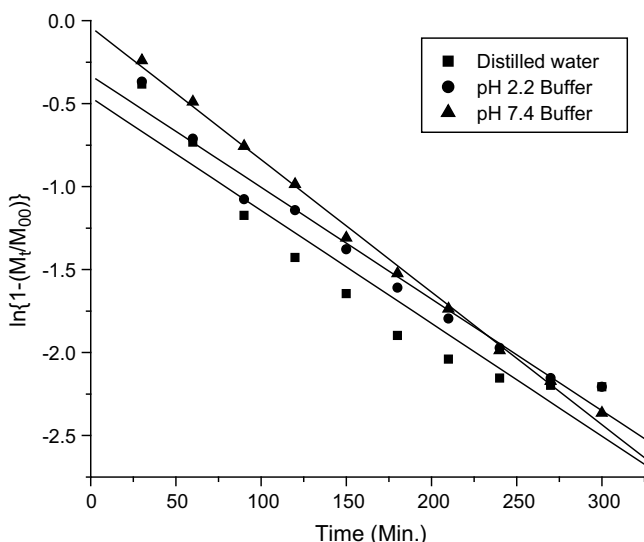
The diffusion exponent ' $n$ ' and gel characteristic constant ' $k$ ' has been obtained from the slope and intercept of the plot of  $\ln M_t/M_\infty$  versus  $\ln t$ , for the release of ranitidine hydrochloride from the hydrogels (Fig. 4.3). The diffusion exponent ' $n$ ' have 0.43, 0.44 and 0.62 values and gel characteristic constant ' $k$ ' have  $8.669 \times 10^{-2}$ ,  $8.016 \times 10^{-2}$  and  $2.964 \times 10^{-2}$  values in distilled water, pH 2.2 buffer and pH 7.4 buffer. It is clear from the values of the ' $n$ ' that the release of drug from the drug-loaded hydrogels occurred through Fickian diffusion mechanism in distilled water and in pH 2.2 buffer and non-Fickian type of diffusion mechanism in pH 7.4 buffer. The values of the diffusion coefficients for the release of drug from the hydrogels in different pH buffer are obtained from the Figs. 4.4 and 4.5, and are presented in Table 2. The values obtained for initial diffusion coefficient are higher than the average and late diffusion coefficient. Higher values of the initial diffusion



**Fig. 4.3.** (Singh and Sharma) Plot for the evaluation of diffusion exponent ' $n$ ' and gel characteristic constant ' $k$ ' for the drug release from sterculia-cl-poly(AAm) hydrogels in different medium at 37 °C.



**Fig. 4.4.** (Singh and Sharma) Plot of  $M_t/M_\infty$  versus  $t$  for the evaluation of initial diffusion coefficient ( $D_i$ ) for the drug release from sterculia-cl-poly(AAm) hydrogels in different medium at 37 °C.



**Fig. 4.5.** (Singh and Sharma) Plot of  $\ln\{1 - (M_t/M_\infty)\}$  versus time for the evaluation of late diffusion coefficient ( $D_L$ ) for the drug release from sterculia-cl-poly(AAm) hydrogels in different medium at 37 °C.

coefficient reflect that initially the rate of diffusion of drug from the polymer was higher than the rate of diffusion of drug from the later stages.

#### 4. Conclusion

It is concluded from the foregone discussion that the composition of the hydrogels and pH of the swelling medium affects the swelling of the modified sterculia based hydrogels. During synthesis of polymer matrix the reaction parameters such as monomer concentration, initiator concentration, amount of sterculia gum and concentration of crosslinker have affected the pore size and network density which have been reflected in the swelling of the hydrogels. It is also concluded from the values of diffusion exponent ' $n$ ' that swelling of the hydrogels and release of drug from the drug-loaded hydrogels occurred through Fickian type diffusion

mechanism in distilled water and in pH 2.2 buffer. On the other hand, release of drug from polymer matrix in pH 7.4 buffer has occurred through non-Fickian type of diffusion mechanism. Further solubility of the drug also affects the release pattern.

#### References

- Agnihotri, S. A., Jawalkar, S. S., & Aminabhavi, T. M. (2006). Controlled release of cephalexin through gellan gum beads: Effect of formulation parameters on entrapment efficiency, size, and drug release. *European Journal of Pharmaceutics and Biopharmaceutics*, 63(3), 249–261.
- Alfrey, T., Gurnee, E. F., & Lloyd, W. G. (1966). Diffusion in glassy polymers. *Journal of Polymer Science. Part C*, 12, 249–261.
- Baveja, S. K., Ranga Rao, K. V., & Arora, J. (1988). Examination of natural gums and mucilages as sustaining materials in tablet dosage forms. *Indian Journal of Pharmaceutics Sciences*, 50, 89–92.
- Behall, K. M. (1990). Effect of soluble fibers on plasma lipids, glucose tolerance and mineral balance. *Advanced Experimental Medicinal Biology*, 270, 7–16.
- Behall, K. M., Scholfield, D. J., Lee, K., Powell, A. S., & Mores, P. B. (1987). Mineral balance in adult men: Effect of four fibers. *American Journal of Clinical Nutrition*, 46(2), 307–314.
- Bhardwaj, T. R., Kanwar, M., Lal, R., & Gupta, A. (2000). Natural gums and modified natural gums as sustained-release carriers. *Drug Development and Industrial Pharmacy*, 26(10), 1025–1038.
- Capron, J. P., Zeitoun, P., & Julien, D. (1981). A multicenter controlled trial of a combination of kaolin, sterculia gum, meprobamate, and magnesium salts, in the irritable bowel syndrome. *Gastroenterology Clinical Biology*, 5(1), 67–72.
- Cascone, M. G., Barbani, N., Cristallini, C., Giusti, P., Ciardelli, G., & Lazzeri, L. (2001). Bioartificial polymeric materials based on polysaccharides. *Journal of Biomaterial Sciences. Polymer Edition*, 12(3), 267–281.
- Chang, C. P., Leung, T. K., Lin, S. M., & Hsu, C. C. (2006). Release properties on gelatin-gum arabic microcapsules containing camphor oil with added polystyrene. *Colloids and Surface. B, Biointerfaces*, 50(2), 136–140.
- Chaurasia, M., Chourasia, M. K., Jain, N. K., Jain, A., Soni, V., Gupta, Y., et al. (2006). Cross-linked guar gum microspheres: A viable approach for improved delivery of anticancer drugs for the treatment of colorectal cancer. *AAPS Pharmaceutical Sciences and Technology*, 7(3), E1–E9.
- Chen, J., Jo, S., & Park, K. (1995). Polysaccharide hydrogels for protein drug delivery. *Carbohydrate Polymers*, 28, 69–76.
- Coviello, T., Alhaique, F., Dorigo, A., Matricardi, P., & Grassi, M. (2007). Two galactomannans and scleroglucan as matrices for drug delivery: Preparation and release studies. *European Journal of Pharmaceutics and Biopharmaceutics*, 66(2), 200–209.
- Das, A., Wadhwa, S., & Srivastava, A. K. (2006). Cross-linked guar gum hydrogel discs for colon-specific delivery of ibuprofen: Formulation and in vitro evaluation. *Drug Delivery*, 13(2), 139–142.
- Durso, D. F. (1980). In R. Davidson (Ed.), *Handbook of water soluble gums and resins* (pp. 1–2). New York: McGraw Hill, Kingsport Press.
- Gauthami, S., & Bhat, V. R. (1992). *A monograph on gum karaya*. Hyderabad: National Institute of Nutrition, Indian Council of Medical Research.
- George, M., & Abraham, T. E. (2007). pH sensitive alginate-guar gum hydrogel for the controlled delivery of protein drugs. *International Journal of Pharmaceutics*, 335(1–2), 123–129.
- Guerre, J., & Neuman, M. (1979). Treatment of chronic colonic diseases with a new topical digestive agent, mucilage (karaya gum) combined with polyvinylpyrrolidone (P.V.P.P.). *Médecine & Chirurgie Digestives*, 8(7), 679–682.
- Hunold, W. (1979). Functional and organic intestinal diseases. Therapeutic experience using Karaya bismuth. *ZFA (Stuttgart)*, 55(15), 951–955.
- Huttel, E. (1983). Treatment of acute diarrhea in general practice. Therapeutic experiences with karaya bismuth. *Die Medizinische Welt*, 34(48), 1383–1384.
- Ji, C., Xu, H., & Wu, W. (2007). In vitro evaluation and pharmacokinetics in dogs of guar gum and Eudragit FS30D-coated colon-targeted pellets of indomethacin. *Journal of Drug Target*, 15(2), 123–131.
- Kanamori, Y., Hashizume, K., Sugiyama, M., Tomonaga, T., Takayasu, H., Ishimaru, T., et al. (2007). Long-term survival of a baby with body stalk anomaly: Report of a case. *Surgical Today*, 37(1), 30–33.
- Korsmeyer, R. W., & Peppas, N. A. (1981). Effect of morphology of hydrophilic polymer matrices on the diffusion and release of water-soluble drugs. *Journal of Membrane Science*, 9, 211–227.
- Krishnaiah, Y. S. R., Muzib, Y. I., Rao, G. S., Bhaskar, P., & Satyanarayana, V. (2003). Studies on the development of colon targeted oral drug delivery systems for ornidazole in the treatment of amoebiasis. *Drug Delivery*, 10, 111–117.
- Leung, A. Y. (1980). *Encyclopedia of common natural ingredients used in food, drugs, and cosmetics*. New York, NY: John Wiley and Sons.
- Love-Mignogna, S., & Wind, S. (1978). Decubitus ulcers and the karaya treatment program. *ONA Journal*, 5(9), 17–18.
- May, C. B. (1982). Karaya gel in the treatment of leg ulcers. *Nursing Times*, 78(29), 1233–1234.
- Meier, P., Seiler, W. O., & Stahelin, H. B. (1990). Bulk-forming agents as laxatives in geriatric patients. *Schweizerische Medizinische Wochenschrift*, 120, 314–317.
- Mostafa, K. M., & Morsy, M. S. (2004). Modification of carbohydrate polymers via grafting of methacrylonitrile onto pregelated starch using potassium monopersulfate/Fe<sup>2+</sup> redox pair. *Polymer International*, 53(7), 885–890.



- Mundargi, R. C., Patil, S. A., Agnihotri, S. A., & Aminabhavi, T. M. (2007). Development of polysaccharide-based colon targeted drug delivery systems for the treatment of amoebiasis. *Drug Development and Industrial Pharmacy*, 33(3), 255–264.
- Mundargi, R. C., Patil, S. A., & Aminabhavi, T. M. (2007). Evaluation of acrylamide-grafted-xanthan gum copolymer matrix tablets for oral controlled delivery of antihypertensive drugs. *Carbohydrate Polymers*, 69, 130–141.
- Munday, D. L., & Cox, P. J. (2000). Compressed xanthan and karaya gum matrices: Hydration, erosion and drug release mechanisms. *International Journal of Pharmaceutics*, 203(1–2), 179–192.
- Nishimura, S., Miyura, Y., Ren, L., Sato, M., Yamagishi, A., Nishi, N., et al. (1993). An efficient method for the synthesis of novel amphiphilic polysaccharide by region and thermo selective modifications of chitosan. *Chemistry letters*, 1623–1626.
- Park, C. R., & Munday, D. L. (2004). Evaluation of selected polysaccharide excipients in buccoadhesive tablets for sustained release of nicotine. *Drug Development and Industrial Pharmacy*, 30(6), 609–617.
- Patel, V. F., & Patel, N. M. (2007). Statistical evaluation of influence of xanthan gum and gaur gum blends on dipyridamole release from floating matrix tablets. *Drug Development and Industrial Pharmacy*, 33(3), 327–334.
- Peppas, N. A., & Korsmeyer, R. W. (1987). Dynamically swelling hydrogels in controlled release applications, in hydrogels, in medicines and pharmacy. In N. A. Peppas (Ed.), *Properties and Applications* (Vol. III, pp. 118–121). Boca Raton, FL: CRC Press Inc..
- Pharmacopoeia of India, (1985), (3rd ed.), Vol. II, Controller of publications, Delhi, Appendix-7, pp. A-142.
- Ray, A. R., Batra, V., Bhowmick, A., & Behra, B. K. (1994). Sustained release of ferrous sulfate from polymer-coated gum arabica pellets. *Journal of Pharmaceutical Sciences*, 83, 632–635.
- Ritger, P. L., & Peppas, N. A. (1987a). A simple equation for description of solute release I. Fickian and non-Fickian release from non-swellable devices in the form of slabs, spheres, cylinders or discs. *Journal of Controlled Release*, 5, 23–36.
- Ritger, P. L., & Peppas, N. A. (1987b). A simple equation for description of solute release I. Fickian and non-Fickian release from swellable devices. *Journal of Controlled Release*, 5, 37–42.
- Singh, B. (2007). Psyllium as therapeutic and drug delivery agent. *International Journal Pharmaceutics*, 334, 1–14.
- Verbeken, D., Dierckx, S., & Dewettinck, K. (2003). Exudate gums: Occurrence, production, and applications. *Applied Microbiology and Biotechnology*, 63(1), 10–21.
- Verhoeven, E., Vervaeke, C., & Remon, J. P. (2006). Xanthan gum to tailor drug release of sustained-release ethylcellulose mini-matrices prepared via hot-melt extrusion: In vitro and in vivo evaluation. *European Journal Pharmaceutics and Biopharmaceutics*, 63(3), 320–330.
- Weiping, W. (2000). Tragacanth and karaya. In G. O. Philips & P. A. Williams (Eds.), *Handbook of hydrocolloids* (pp. 155–168). Cambridge: Woodhead.
- Wilson, M., & Harvey, W. (1989). Prevention of bacterial adhesion to denture acrylic. *Journal of Dentistry*, 17(4), 166–170.
- Zhang, L. M., Wang, G. H., Lu, H. W., Yang, C., & Yan, L. (2005). A new class of starch-based hydrogels incorporating acrylamide and vinyl pyrrolidone: Effects of reaction variables on water sorption behavior. *Bioactive and Compatible Polymers*, 20(5), 491–501.
- Zide, B. M., & Bevin, A. G. (1980). Treatment of shallow soft tissue ulcers with an infrequent dressing technique. *Anal Plastic Surgery*, 4(1), 79–83.
- Zohuriaan-Mehr, M. J., Motazedi, Z., Kabiri, K., & Ershad-Langroudi, A. (2005). New super-absorbing hydrogel hybrids from gum arabic and acrylic monomers. *Journal of Macromolecular. Science. Part A: Pure and Applied Chemistry*, 42, 1655–1666.